



A Geno Technology, Inc. (USA) brand name

# **Protein A Magnetic Beads**

(Cat. # 786-902, 786-903)



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#### INTRODUCTION

G-Biosciences' Protein A Magnetic Beads are designed as a rapid and simple tool for immunoprecipitation, purification and/or depletion assays, and other magnetic separation applications. Antibodies easily bind to the Protein A Magnetic Beads due to their high affinity for protein A. The target protein is captured by the antibody:Protein A Magnetic Beads and are temporarily immobilized at the tube wall by magnetic attraction and all other proteins are easily removed.

#### ITEMS SUPPLIED

Cat. #	Description	Size
786-902	Protein A Magnetic Beads	1ml
786-903	Protein A Magnetic Beads	5 x 1ml

# STORAGE CONDITIONS

The beads are shipped at ambient temperature. Upon arrival, store the beads at 4°C. If stored and handled correctly the beads have a 1 year shelf life.

#### **SPECIFICATIONS**

 $Fe_3O_4$  beads coated with dextran of an average 1 $\mu$ m in diameter. Protein A is coupled covalently to dextran. Supplied in phosphate buffered saline, pH 7.4, containing 0.02% Tween 20 and 0.09% sodium azide.

#### **PRECAUTIONS**

- Do not freeze the magnetic beads
- Do not store near magnetic sources

# **BINDING PROPERTIES**

The binding capacity of Protein A Magnetic Beads is more than  $26\mu g$  human IgG per  $100\mu l$ . The binding strength of Protein A Magnetic Beads to different immunoglobulins is listed as below

Species	Antibody Class	Protein A	Protein G
Mouse	Total IgG	++++	++++
	IgG <sub>1</sub>	+	+++
	IgG <sub>2a</sub>	++++	++++
	IgG <sub>2b</sub>	++++	++++
	IgG₃	+++	+++
Human	Total IgG	++++	++++
	$IgG_1$	++++	++++
	IgG₂	++++	++++
	IgG₃	+	++++
	IgG <sub>4</sub>	++++	++++
Rat	Total IgG	+	++
	$IgG_1$	-	+
	$lgG_{2a}$	-	++++
	IgG <sub>2b</sub>	-	++
	IgG <sub>2c</sub>	++	+++
Hamster	Total IgG	++	++
Guinea Pig	Total IgG	++++	++
Rabbit	Total IgG	++++	+++
Horse	Total IgG	++	++++
Cow	Total IgG	++	++++
Pig	Total IgG	+++	++
Sheep	Total IgG	+	++
Goat	Total IgG	+	++
Chicken	Total IgG	-	-

Table 1: Relative affinity of Protein A and Protein G for Immunoglobulins

# ADDITIONAL ITEMS REQUIRED

- Washing Buffer: 1X PBS buffer (pH 7.4) with 0.02% Tween 20
- Elution Buffer: 100mM glycine, pH 3.0
   Neutralization Buffer: 1M Tris-HCl, pH 8.5
- Storage Buffer: 0.01M NaH<sub>2</sub>PO<sub>4</sub>, 0.15M NaCl, 2.7mM KCl, pH 7.4, 20% ethanol
- Magnetic Stand or magnet

#### PREPARATION BEFORE USE

Sample preparation: We recommend that for optimal binding the serum samples/ascites fluid or tissue culture media be the addition of 1/10<sup>th</sup> volume of 1.0M Tris, pH 8.0.

# **PROTOCOL**

# Preparation of Protein A Magnetic Beads

- Resuspend the Protein A Magnetic Beads thoroughly by pipetting or vortexing the vial for 15 seconds.
- 2. Transfer adequate amount of Protein A Magnetic Beads into a clean tube.
  NOTE: Take appropriate amount of beads according to the binding capacity mentioned above. For example, if 2-3µg of antibody is used in immunoprecipitation test, 5-10µl of Protein A Magnetic Beads is enough for one test. Excess amount of beads may cause high background in some cases.
- 3. Place the tube on the magnetic stand for 30-60 seconds to immobilize the beads at tube wall.
- 4. Discard the supernatant by aspiration with a pipette.
- 5. Remove the tube from the magnetic stand.
- 6. Add 200µl Washing Buffer and resuspend the beads by pipetting.
- Place the tube on the magnetic stand for 30-60 seconds to immobilize the beads at tube wall.
- 8. Discard the supernatant, and then remove the tube from the magnetic stand.
- 9. Repeat steps 6-8 twice.

**NOTE:** Protein A Magnetic Beads contains 0.09% NaN<sub>3</sub>, so we strongly recommend washing the beads at least three times before use.

# Binding of Antibody

- 10. Mix appropriate amount of antibody in  $200\mu l$  Washing Buffer and transfer to the tube from step 9 and vortex for 10 seconds.
- 11. Incubate for 30 minutes at room temperature with gentle rocking or shaking.
- 12. Place the tube on the magnetic stand for 30-60 seconds to immobilize the beads at tube wall.
- 13. Discard the supernatant, and then remove the tube from the magnetic stand.
- 14. Add 200µl Washing Buffer and resuspend the beads by pipetting.

- 15. Place the tube on the magnetic stand for 30-60 seconds to immobilize the beads at tube wall.
- 16. Discard the supernatant, and then remove the tube from the magnetic stand.
- 17. Repeat steps 14-16 two more times to remove unbound antibody.

#### IMMUNOPRECIPITATION METHOD

- 1. Add at least  $100\mu$ l cell lysate sample containing target antigen to the tube from step 17 and vortex for 10 seconds.
- Incubate for 30 minutes at room temperature or 4°C overnight with gentle rocking or shaking.
- Place the tube on the magnetic stand for 30-60 seconds to immobilize the beads at tube wall.
- 4. Discard the supernatant, and then remove the tube from the magnetic stand.
- 5. Add 200µl Washing Buffer and resuspend the beads by pipetting.
- Place the tube on the magnetic stand for 30-60 seconds to immobilize the beads at tube wall.
- 7. Discard the supernatant, and then remove the tube from the magnetic stand.
- 8. Repeat steps 5-7 two more times to remove unbound antigen.
- For SDS-PAGE and Western blot analysis, add appropriate volume of SDS-PAGE
   Loading Buffer and heat the beads at 95°C for 5 minutes. Load the bead/ loading
   buffer mix directly onto an SDS-PAGE gel and proceed with electrophoresis and
   blotting as normal.

# **ELUTION OF ANTIBODY & ANTIGEN**

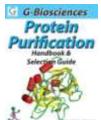
- Add 20µl Elution Buffer to the bead:antibody:antigen mix and resuspend by pipetting.
- 2. Incubate with tilt rotation for 2 minutes at room temperature.
- 3. Place the tube on the magnet stand for 30-60 seconds.
- 4. Collect the supernatant to a clean tube, and then adjust the pH by adding  $2\mu$ l Neutralization Buffer (e.g. 1M Tris-HCl, pH 8.5).

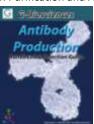
# **TROUBLESHOOTING**

Issue	Suggested Reason	Possible Solution
	Improper resuspension & mixing	Make sure the beads are suspended thoroughly before use Mix beads and sample thoroughly and continuously with either a tilt rotation device or a vortexer.
Immunoglobin binding is low.	Protein A has low binding affinity for antibody	Refer to Table 1 to match the binding preference of protein A with various Immunoglobins
	Incubation time and temperature not ideal	Incubation time and temperature can be optimized depending on the sample column and affinity of antibody for target antigen
Non-specific and background binding is high.	Excess of beads used	Reduce the usage amount of beads per test according to the binding capacity
	Insufficient washing	After incubate bead:antibody complex with antigen, increase wash procedures to >5 times.
	Strong non-specific interactions	Increase the concentration of Tween 20 to 0.1% in Washing Buffer prior to elute the sample
	Cross contamination	Ensure all the Washing Buffer is removed

# RELATED PRODUCTS

Download our Protein Purification and Antibody Production Handbooks.





http://info.gbiosciences.com/complete-protein-purification-handbook/http://info.gbiosciences.com/complete-Antibody-Production-handbook/

For other related products, visit our website at www.GBiosciences.com or contact us.

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